SUPPLEMENT

Biología Tropical

https://doi.org/10.15517/rev.biol.trop..v71iS1.54879

Nursery-reared coral outplanting success in an upwelling-influenced area in Costa Rica

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Received 27-VIII-2022. Corrected 09-II-2023. Accepted 14-II-2023.

ABSTRACT

Introduction: Environmental and intrinsic factors such as seawater temperature, salinity, nutrient concentration, upwelling, species, and life history can influence coral outplant survival and growth, and in consequence, the effectiveness of restoration. Thus, it is key to understand how these factors can shape coral outplant performance to ensure the long-term success of a restoration program.

Objective: To establish the survival and growth rate of outplanted coral nursery-reared colonies of branching *Pocillopora* spp. and massive corals *Pavona gigantea*, *Pavona clavus*, and *Porites lobata* in Bahía Culebra, North Pacific of Costa Rica, and to determine whether the site of origin of the coral fragment and the presence of seasonal upwelling affected the growth of *Pocillopora* outplants.

Methods: From September 2020 to September 2021, we monitored the survival, health, and growth of 30 *Pocillopora* spp. colonies from six donor sites, and 31 fragments of massive species (*P. gigantea* [n = 18], *P. clavus* [n = 8], *P. lobata* [n = 5]) that were outplanted to a degraded reef. We recorded *in situ* seawater temperature, salinity, and nutrient concentration.

Results: By the end of the year, 100 % of the *Pocillopora* spp. outplants survived. Survival was 71.4 % for *P. clavus*, 47.5 % for *P. gigantea*, and 20 % for *P. lobata*. Coral tissue loss and predation marks were observed at least once in 71 % of massive outplants. *Pocillopora* spp. colonies increased their initial area in 333.1 %, at a rate of 9.98 ± 1.69 cm yr⁻¹. The site of origin of the outplant influenced growth rate, but not the presence of seasonal upwelling. Massive species fragments grew at a rate of 1.35 ± 0.24 cm yr⁻¹ for *P. clavus*, 1.48 ± 0.21 cm yr⁻¹ for *P. gigantea*, and 0.61 cm yr⁻¹ for *P. lobata*, with no differences among them.

Conclusions: Environmental conditions at site level allow for high survival and growth of *Pocillopora* spp. outplants, previously considered as sensitive to stressors, and point towards acclimation to seasonal upwelling. Although survival of massive species was lower, developing a multi-species approach is key to enhance restoration success.

Key words: Bahía Culebra; coral reef; coral gardening; Eastern Tropical Pacific; ecological restoration; growth rate.

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RESUMEN

Éxito de trasplante de colonias de coral cultivadas en vivero en un área influenciada por afloramiento en Costa Rica.

Introducción: Factores ambientales e intrínsecos como la temperatura del agua, salinidad, concentración de nutrientes, afloramiento, especies e historia de vida pueden influir en la supervivencia y crecimiento de los trasplantes de coral y, en consecuencia, en la eficacia de la restauración. Por ello, entender cómo estos factores pueden moldear el desempeño de los trasplantes es clave para asegurar el éxito de un programa de restauración a largo plazo.

Objetivo: Establecer la supervivencia y tasa de crecimiento de los trasplantes de colonias de coral previamente cultivadas en viveros del coral ramificado *Pocillopora* spp. y las especies de crecimiento masivo *Pavona gigantea, Pavona clavus* y *Porites lobata* en Bahía Culebra, Pacífico Norte de Costa Rica, y determinar si el sitio donante de origen del fragmento de coral y la presencia de afloramiento estacional afectó el crecimiento de los trasplantes de *Pocillopora*.

Métodos: De setiembre 2020 a setiembre 2021, se monitoreó la supervivencia, salud y crecimiento de 30 colonias de *Pocillopora* spp. de seis sitios donantes diferentes y 31 fragmentos de especies masivas (*P. gigantea* [n = 18], *P. clavus* [n = 8], *P. lobata* [n = 5]) que se trasplantaron a un arrecife degradado. Se registró la temperatura del agua, la salinidad y concentración de nutrientes *in situ*.

Resultados: Al final del año, el 100 % de los trasplantes de *Pocillopora* spp. sobrevivieron. La supervivencia fue de 71.4 % para *P. clavus*, 47.5 % para *P. gigantea* y 20 % para *P. lobata*. Se observaron pérdidas de tejido y marcas de depredación al menos una vez en un 71 % de los trasplantes masivos. Las colonias de *Pocillopora* spp. aumentaron su área inicial en un 333.1 %, a una tasa de 9.98 ± 1.69 cm año⁻¹. El sitio de origen del trasplante tuvo efecto sobre la tasa de crecimiento, pero no la presencia de afloramiento estacional. Los fragmentos de especies masivas crecieron a tasas de 1.35 ± 0.24 cm año⁻¹ (*P. clavus*), 1.48 ± cm año⁻¹ (*P. gigantea*) y 0.61 cm año (*P. lobata*), sin diferencias entre ellas.

Conclusiones: Las condiciones del sitio permiten una alta supervivencia y crecimiento de trasplantes de *Pocillopora* spp., que previamente habían sido considerados sensibles a factores estresantes, e indican aclimatación a las condiciones locales de afloramiento estacional. Pese a que la supervivencia de las especies masivas fue menor, es esencial desarrollar un enfoque multi-especie para aumentar el éxito de la restauración.

Palabras clave: Bahía Culebra; arrecife coralino; jardinería de coral; Pacífico Tropical Oriental; restauración ecológica; tasa de crecimiento.

INTRODUCTION

As coral reefs worldwide are declining, active solutions have been developed over the last few decades in order to reverse this alarming trend and restore the function of these ecosystems (Hughes et al., 2019; Lirman & Schopmeyer, 2016; Unsworth et al., 2020). Coral gardening is a popular technique to reestablish coral populations and increase coral cover (Ladd et al., 2018; Rinkevich, 1995); this strategy uses fragments from wild donor colonies and grows them in nurseries, followed by their outplanting to a degraded reef (Rinkevich, 2014). Even though the nursery stage of coral restoration and propagation methodologies are well-established, outplanting success can vary, with an average survival of 66 % of

the outplanted corals (Boström-Einarsson et al., 2020), but with some projects reporting a higher survival of up to 89 % (Ware et al., 2020). However, the time frame considered to monitor outplant survival is often short, as most projects do so during a 12-month period (Boström-Einarsson et al., 2020). Coral outplants can undergo rapid mortality due to several stressors, both abiotic and biotic, such as temperature, disease, predation, and presence of competitor species (Cano et al., 2021; Foo & Asner, 2020; Hughes et al., 2017; Koval et al., 2020; Miller et al., 2014; van Woesik et al., 2017; Ware et al., 2020). Hence, there is increasing interest in developing techniques that could improve the efficiency and longterm success of such efforts (Lohr et al., 2020).

Several techniques exist for outplanting coral colonies or fragments onto either suitable natural reef substrate or artificial structures (Ferse et al., 2021). Most restoration practitioners use marine epoxy, cable ties, steel cables or nails to attach corals onto the substrate (Boström-Einarsson et al., 2020; Goergen & Gilliam, 2018; Omori, 2019). Which technique is the most appropriate is highly site-specific, depending on the type of substrate, currents, wave action, herbivore presence, and even colony size and density (Goergen & Gilliam, 2018; van Woesik et al., 2017). Thus, these considerations should be taken into account for the outplanting design before scaling up outplanting efforts.

Therefore, the success of the outplanting stage will also be inevitably affected by the appropriate selection of the outplanting site, which must gather conditions that promote coral survival and growth (see Kleypas et al., 1999). These include environmental factors such as seawater temperature (Coles & Jokiel, 1978; Foo & Asner, 2020), pH (Kleypas & Yates, 2009), nutrients, salinity (D'Angelo & Wiedenmann, 2014; Faxneld et al., 2010; Wiedenmann et al., 2013), and light (Coles & Jokiel, 1978; Foo & Asner, 2021); and biological factors like the presence of associated organisms such as fish, invertebrates, and other benthic groups (Cano et al., 2021; Rivas et al., 2021). Furthermore, intrinsic factors such as the coral species, life history, acclimation to local conditions, and genotype have an equally important role in shaping restoration outcomes (Baums et al., 2019; Goergen & Gilliam, 2018; Lirman et al., 2014).

Once transplanted, monitoring coral outplants is key for measuring outplanting success and thus, evaluating restoration efforts (Edwards, 2010; Foo & Asner, 2020). Despite the lack of standardised monitoring guidelines, outplant survival and growth rate are some of the most widely used assessments for this purpose (Hein et al., 2017; Edmunds & Putnam, 2020).

In the Eastern Tropical Pacific (ETP), coral restoration experiences are limited compared

to other regions such as the Indo-Pacific and Caribbean region (Bayraktarov et al., 2020). In recent years, however, restoration initiatives in the ETP are rapidly increasing, and most of them focus on the branching coral genus Pocillopora (Chomitz, 2021; Combillet et al., 2022; Ishida-Castañeda et al., 2020; Liñán-Cabello et al., 2011; Tortolero-Langarica et al., 2014; Tortolero-Langarica et al., 2019). Direct transplantation in the Central Mexican Pacific showed a survival rate between 89-95.5 % over 270 days (Liñán-Cabello et al., 2011), which go in concordance with results in Colombia, which show 100 % survival after 5-months using nursery-reared Pocillopora fragments (Ishida-Castañeda et al. 2020), with higher growth rates than direct transplants. Outplanting using micro-fragmentation has only been implemented once in the ETP (Tortolero-Langarica et al., 2020), but never in Costa Rica, where only two studies have focused on Pocillopora spp. outplanting success; Guzmán (1991) reported a 79-83 % survival in Caño Island, while Chomitz (2021) reports 83 % survival and an average 68 % growth in area along eight months of study in Golfo Dulce, both in the South Pacific of the country. However, this data cannot be extrapolated as environmental conditions between areas of the Pacific coast of Costa Rica are different, as are reef dynamics and the main coral reef-building species, mainly due to the presence of a seasonal upwelling and lower precipitation in the North Pacific of the country (Cortés et al., 2010).

This study represents the assessment of the first restoration project in the area, in order to evaluate the outplanting success of the first nursery-reared corals outplanted onto a degraded reef, therefore we aimed to (1) monitor the health and survival of outplanted colony fragments of the branching *Pocillopora* spp. and massive *Pavona gigantea* (Verrill, 1869), *Pavona clavus* (Dana, 1846) and *Porites lobata* (Dana, 1846) corals in Bahía Culebra, North Pacific of Costa Rica, (2) quantify the growth rate of outplanted corals in terms of area and diameter and (3) determine whether the origin (donor colonies sites) of the coral fragment and



presence of upwelling have an effect on coral growth for *Pocillopora* spp. colonies, one year post-outplanting.

MATERIALS & METHODS

Study area: Bahía Culebra $(10^{\circ}37'N - 85^{\circ}39'W)$ is a semi-enclosed bay in the Gulf of Papagayo, North Pacific of Costa Rica (Fig. 1). The bay extends for more than 20 km², and reaches its maximum depth at about 42 m (Rodríguez-Sáenz & Rodríguez-Fonseca, 2004). It is influenced by seasonal upwelling from December to April, causing a decrease in seawater temperature up to 10 °C from the annual mean $(27 \pm 0.1 \text{ °C})$ (Jiménez, 2001; Jiménez et al., 2010), and bringing up more acidic (pH 7.8) nutrient-rich waters (Rixen et al., 2012; Sánchez-Noguera et al., 2018b; Stuhldreier et al., 2015).

Bahía Culebra once harboured some of the most extensive coral reefs on the Pacific coast

of the country, mainly built by the branching coral Pocillopora, with high live coral cover $(44.0 \pm 0.3 \%)$ (Jiménez 2001; Cortés & Jiménez, 2003). However, in the last two decades, coral reefs in the bay experienced intense coral bleaching and mass coral mortality (Sánchez-Noguera, 2012) caused by El Niño Southern-Oscillations (ENSO) events, increase in anthropic eutrophication and frequent harmful algal blooms (Fernández, 2007; Jiménez, 2007; Sánchez-Noguera, 2012), resulting in an abrupt loss of coral cover. This situation favoured the recruitment and proliferation of macroalgae (Alvarado et al., 2018; Fernández-García et al., 2012), and a cascading effect on high densities of the sea urchin Diadema mexicanum (A. Agassiz, 1863). As a consequence of sea urchin bioerosion, the coral framework weakened and was undermined (Alvarado et al., 2012; Alvarado et al., 2016). In order to mitigate the damage, a coral restoration project was initiated in Bahía Culebra in September 2019, by using



Fig. 1. Location of Güiri-Güiri outplanting site (yellow square), Jícaro nursery site (red star), and donor colonies sites (blue dot) in Bahía Culebra, North Pacific of Costa Rica.

coral gardening techniques and a subsequent transplant to nearby sites in the bay (Fabregat-Malé et al., *in prep.*).

Experimental design: The species of corals used in this study were the branching Pocillopora genus, and the massive P. gigantea, P. clavus, and P. lobata. Here, we worked with Pocillopora damicornis (Linnaeus, 1758) and Pocillopora elegans (Dana, 1846). To visually distinguish these species based on their morphology is equivocal (Pinzón et al., 2013); thus, considering their functional and genetic similarities (Pinzón & LaJeunesse, 2011; Pinzón et al., 2013), they were grouped and further referred as "Pocillopora spp.". Coral fragments had been previously obtained from donor colonies on different reefs and coral communities around Bahía Culebra (Fig. 1). Donor colonies were randomly selected at depths between 2-7 m, growing at least 5 m from each other so as to maximize chances of sampling distinct genets. For Pocillopora spp., five fragments of about 2-5 cm were originally extracted from each donor colony, while for massive species (P. clavus, P. gigantea and P. lobata), microfragments of around 1.5-2 cm² were obtained, using a diamond band saw (Model C-40, Gryphon, USA) (Fabregat-Malé et al., in prep.).

Nursery-reared coral colonies were outplanted in September 2020 to Güiri-Güiri reef (10° 36' 49.7" N, 85° 41' 24.1" W), in Bahía Culebra (Fig. 1), after a one-year cultivation in the nurseries. The reef formation in Güiri-Güiri is mainly built by dead *Pavona* framework, and some sparse *Pocillopora*, *P. gigantea* and *P. clavus* colonies (Jiménez, 1997; Jiménez, 2007). Coral colonies were outplanted at 5 m depth and on dead coral substrate, comprising an area of approximately 100 m² on the backreef area.

A total of 30 Pocillopora spp. colonies were tagged and outplanted onto the dead coral substrate using steel nails and cable ties (Fig. 2A), with a minimum distance of 50 cm from each other. Nursery-reared Pocillopora spp. colonies originally came from six different donor sites on the bay (Fig. 1): Matapalo (n = 11), Sombrero (n = 7), Esmeralda (n = 7)4), Jícaro (n = 4), Güiri-Güiri (n = 3), Palmitas (n = 1). For the massive coral species, an underwater drill (Nemo V2, NemoPowerTools, USA) was used to outplant 31 fragments (n =18 P. gigantea fragments, n = 8 P. clavus, and n = 5 P. lobata), which were then fixed to the substrate using marine epoxy glue (Marine-Tex, USA) (Fig. 2B). The small sample sizes of massive species used during outplanting are due to their low survival in the nursery stage (Fabregat-Malé et al., in prep.). Massive fragments from the same species and genotype (i.e., donor colony) were outplanted together (2 cm apart) so as to promote fusion when growing (Forsman et al., 2015).



Fig. 2. A. Outplanting technique for nursery-reared *Pocillopora* spp. colonies and B. for massive species (*Porites lobata*, *Pavona gigantea* and *Pavona clavus*) fragments, in Güiri-Güiri reef, Costa Rica.



Monitoring occurred monthly postoutplanting for 12 months, from September 2020 to September 2021. The survival of each individual colony and fragment (alive, pale/ bleached, dead or lost) was recorded, as well as partial tissue mortality. Coral growth was also recorded monthly, establishing a photoquadrat around each colony using a polyvinyl chloride (PVC) frame (30x30 cm) that held the camera above the substrate and the coral colonies. Photographs were taken from the same angle and direction, with a scaling object, using a Nikon COOLPIX W300 underwater camera. Each image was analysed using the photo-analysis software PhotoQuad (Trygonis & Sini, 2012), in order to calculate the area of outplanted coral colonies (cm²) and colony diameter (cm), evaluating lateral growth of colonies. Additionally, sea urchin D. mexicanum density was also censed, using belt-transects 10 x 2 m (20 m²) along the outplanting area, and used as an indicator of bioerosion and potential population controller of other groups (i.e., macroalgae and turf) through herbivory (Alvarado et al., 2012).

To determine the environmental conditions to which coral outplants were subject, seawater temperature at site was recorded every 30 min using HOBO[®] data loggers. Six water samples were collected *in situ* monthly to determine salinity (PSU) and nutrient concentration $(NO_3^-, NO_2^-, PO_4^{3-}, NH_4^+, and SiO_4)$, using a plastic syringe with a 1 µm glass microfiber filter, and were later processed using a field refractometer (PCE-0100, PCE Instruments, Germany) and a continuous flow autoanalyzer (QuikChem 8500, Lachat Instruments, USA), respectively.

Data analysis: Survival curves between species were compared using a Kolmogorov-Smirnov (KS) test. Lost fragments were considered as "dead" for calculating survival rates. Differences in outplant size from initial to final month were analysed using a Student's t-test. Growth rate of coral outplants was calculated over the first year after transplantation; the area (cm²) and diameter (cm) of each Pocillopora spp. colony and massive fragment were subtracted from the measurement of the following month. Growth rate results from colonies and fragments that had survived throughout monitoring period. Pocillopora spp. growth rates (cm mo⁻¹) between upwelling and nonupwelling season were compared using a Student's t-test. Differences in Pocillopora growth rate (cm² mo⁻¹) between sites of origin of



Fig. 3. Cumulative survivorship (%) of massive coral fragments (*Pavona clavus* n = 8, in red; *Pavona gigantea* n = 18, in yellow; and *Porites lobata* n = 5, in blue) in the outplanting site of Güiri-Güiri, Costa Rica, through monitoring period (September 2020-September 2021).

donor colonies were analyzed using a one-way ANOVA, followed by Tukey HSD post-hoc tests. Palmitas was excluded as a donor site from this analysis as only one colony from this site was outplanted. Growth rates between massive species were also compared using a non-parametric Kruskal-Wallis test. Data were tested for normality and equality of variances using the Shapiro-Wilk test and Bartlett test, respectively, to ensure they met model assumptions. Statistical analyses were performed using R (R Core Team, 2021).

RESULTS

Outplant survival: Exactly one year after their transplantation to the Güiri-Güiri reef, 100 % of the *Pocillopora* spp. outplants survived, none were lost, and no bleaching occurred. When grouping massive species together, overall survival by the end of the first year was 48.4 % (*P. clavus* = 71.4 %, *P. gigantea* = 47.5 %, and *P. lobata* = 20.0 %) (Fig. 3). Decrease in massive species survival was a result of the loss of fragments during monitoring period, not their death. This decline in survival in massive species was most pronounced in August 2021. Survival curves significantly differed only between *P. clavus* and *P. lobata* (D = 0.727, P < 0.05). Partial mortality and predation marks were only observed in outplants of massive species, with 22 fragments (71 %) having experienced partial tissue loss at least once during monitoring period (Fig. 4). Paleness in *P. lobata* fragments was observed in December 2020, but fragments recovered their coloration the following month.

Outplant growth: *Pocillopora* spp. colonies were initially $63.24 \pm 36.48 \text{ cm}^2$ ($8.66 \pm 2.38 \text{ cm}$ in diameter). One year after outplanting, colonies significantly grew to $273.91 \pm 105.81 \text{ cm}^2$ and $18.64 \pm 3.46 \text{ cm}$ in diameter ($F_{1,58} = 106.3, \text{P} < 0.005$) (Fig. 5). This represents a 333.1 % increase in area, and a 115.2 % increase in diameter from initial size. The rate of change in area of the *Pocillopora* spp. outplants was calculated to be $17.56 \pm 6.28 \text{ cm}^2 \text{ mo}^{-1}$ (9.98 $\pm 1.69 \text{ cm} \text{ yr}^{-1}$) (Table 1).

Despite the loss, massive species significantly increased their area from initial size in 157.3 % for *P. gigantea* fragments ($R^2 = 0.6421$, $F_{1,147} = 266.6$, P < 0.005), 161.6 % for *P. clavus* ($R^2 = 0.7496$, $F_{1,76} = 231.5$, P < 0.005), and 70.4 % in *P. lobata* ($R^2 = 0.5452$,



Fig. 4. Percentage (%) of outplanted fragments of massive species (*Pavona clavus*, in red; *Pavona gigantea*, in yellow; and *Porites lobata*, in blue) exhibiting partial tissue loss through monitoring period (September 2020 to September 2021) in Güiri-Güiri reef, Costa Rica.

Revista de Biología Tropical, ISSN: 2215-2075 Vol. 71 (S1): e54879, abril 2023 (Publicado Abr. 30, 2023)



Fig. 5. A. Increase of *Pocillopora* spp. colony area (cm²) from September 2020 to September 2021, **B.** and growth of the same *Pocillopora* spp. outplant at the start of monitoring period (left), after six months (middle), and after one year (right), in the outplanting site in Güiri-Güiri reef, Costa Rica.

 TABLE 1

 Initial and final sizes (mean area and diameter ± SD), and mean growth rate (± SD) of outplanted species to Güiri-Güiri restoration site, Costa Rica.

| Species | Initial size | | Final size | | |
|------------------|-------------------------|-----------------|-------------------------|----------------|---|
| | Area (cm ²) | Diameter (cm) | Area (cm ²) | Diameter (cm) | $\frac{1}{m}$ Growin rate (cm ² yr ⁻¹) |
| Pocillopora spp. | 63.24 ± 36.48 | 8.66 ± 2.38 | 273.91 ± 105.81 | 18.64 ± 3.46 | 210.67 ± 75.41 |
| Pavona gigantea | 5.29 ± 1.25 | 2.24 ± 0.28 | 13.61 ± 2.87 | 4.11 ± 0.35 | 8.18 ± 2.29 |
| Pavona clavus | 4.37 ± 0.91 | 2.37 ± 0.49 | 11.43 ± 1.43 | 3.80 ± 0.20 | 6.69 ± 1.63 |
| Porites lobata | 4.60 ± 1.25 | 2.40 ± 0.35 | 7.84 | 3.16 | 2.77 |

 $F_{1,25} = 32.17$, P < 0.005). Growth rates for massive species were 1.35 ± 0.24 cm yr⁻¹ for *P. clavus*, 1.48 ± 0.21 cm yr⁻¹ for *P. gigantea* and 0.61 cm yr⁻¹ for *P. lobata* (Fig. 6, Table 1). Growth rates between massive species did not differ statistically ($F_{2,220} = 2.934$, P = 0.0553).

Monthly average growth rate of *Pocillopora* spp. outplants showed statistical

differences through time, with a post-hoc Tukey test revealing lower growth rates in October 2020 (P = 0.0941).

Outplant growth and environmental conditions: Mean seawater temperature in the outplanting site was 27.75 ± 1.90 °C, with a minimum of 19.08 °C and a maximum of 30.52 °C (Appendix 1). Significant differences in

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Fig. 6. Changes in area (cm²) of outplanted fragments of massive fragments (*Pavona clavus*, in red; *Pavona gigantea*, in yellow; and *Porites lobata*, in blue) over monitoring period (September 2020-September 2021) in Güiri-Güiri reef, Costa Rica.

TABLE 2 Nutrient concentration (µM) in outplanting site Güiri-Güiri, Costa Rica, from September 2020 to September 2021.

| Nutrient | Mean (± SD) | Maximum | Minimum | Upwelling season (mean ± SD) | Non-upwelling season (mean ± SD) |
|--|---------------|---------|---------|---------------------------------|-------------------------------------|
| Nitrate (NO ₃ ⁻) | 3.82 ± 2.06 | 7.23 | 1.30 | $2.51 \pm 0.33*$ | $4.95 \pm 2.28*$ |
| Nitrite (NO_2^{-}) | 3.03 ± 0.38 | 3.79 | 2.48 | 2.85 ± 0.22 | 3.19 ± 0.43 |
| Phosphate (PO_4^{3-}) | 0.13 ± 0.13 | 0.34 | UDL | 0.16 ± 0.17 | 0.11 ± 0.09 |
| Ammonium (NH ₄ ⁺) | 4.76 ± 1.17 | 6.71 | 2.93 | 5.18 ± 1.26 | 4.40 ± 1.04 |
| Silicate (SiO ₄) | 1.97 ± 2.83 | 7.93 | UDL | 2.02 ± 3.34 | 1.93 ± 2.59 |

Upwelling season: November to April; Non-upwelling season: May to October. UDL = under detection limit (0.48 μ M). * = P < 0.05.

temperature between upwelling $(26.47 \pm 1.87 \, ^{\circ}\text{C})$ and non-upwelling season $(29.00 \pm 0.76 \, ^{\circ}\text{C})$ were detected $(t_{11437} = 117.08, P < 0.0005)$. Mean seawater salinity was $33.18 \pm 1.05 \, \text{PSU}$, and was significantly higher during dry season $(33.72 \pm 0.81 \, \text{PSU})$ than during rainy season $(32.71 \pm 0.72 \, \text{PSU})$ $(t_{369.47} = -12.906, P < 0.005)$ (Appendix 2). Nutrient concentration was highly variable and did not show any pattern (Appendix 3). Nitrate was the only nutrient whose concentration differed between upwelling and non-upwelling season, with higher concentrations during the non-upwelling rainy season $(F_{1.11} = 6.667, P < 0.05, Table 2)$.

Despite the observed environmental differences between upwelling and non-upwelling season, *Pocillopora* spp. growth rate was not affected ($F_{1.358} = 2.955$, P = 0.0865).

Finally, mean density of *Diadema mexi*canum in the outplanting area was 21.71 ± 1.48 ind m⁻².

Influence of sites of origin on Pocillopora spp. outplant growth: Growth rate of Pocillopora spp. coral colonies differs by site of origin ($F_{5,354} = 10.13$, P < 0.005) (Fig. 7A), as Matapalo outplants presented a higher growth rate (23.81 ± 17.56 cm² mo⁻¹, or 285.66 ± 69.82 cm² yr⁻¹) than the rest of the donor sites (overall



Fig. 7. Differences of mean annual growth rate ($cm^2 yr^1$) of *Pocillopora* spp. outplants between different sites of origin of donor colonies (A), and between outplants originally from Matapalo (in red) and the other sites (B) at Güiri-Güiri restoration site, Costa Rica.

mean = $13.94 \pm 9.10 \text{ cm}^2 \text{ mo}^{-1}$, or $167.25 \pm 32.02 \text{ cm}^2 \text{ yr}^{-1}$) (P < 0.05) (Fig. 7B).

DISCUSSION

Identifying the causes of low survivorship in coral outplants is crucial to improve restoration techniques and their long-term effectiveness (Liñán-Cabello et al., 2011; Ware et al., 2020). In order for coral outplants to become a resource for other organisms, reproduce through fragmentation or sexual mechanisms, and contribute to reef complexity until no further transplantation and/or individual monitoring of the colonies is needed, it is important that they survive during the first year after transplantation and keep growing (Goergen & Gilliam, 2018). While all the outplanted Pocillopora spp. colonies survived during the first year after transplantation, over 50 % of the massive outplants were lost and 67.7 % of the remaining fragments experienced partial mortality and tissue loss, which is common for species with massive growth (Rivas et al., 2021) Different factors could explain the low survivorship in our results: (i) the process of

microfragmentation may damage healthy tissue, in addition to their smaller sizes, which increase their vulnerability to other stressors (Arnold et al., 2010; Forsman et al., 2015), and may result into partial tissue mortality. Nonetheless, microfragments grow at a faster rate than larger corals (Page et al., 2018) and isogenic fusion of fragments may be used for colonies to reach a larger size in a shorter period of time (Forsman et al., 2015); and (ii) the technique implemented, due to the ceramic disk being mechanically dislodged by currents or the interaction of other reef species (Shafir & Rinkevich, 2013; van Woesik et al., 2017), including the presence of herbivores or predators. Large and dense patches of the sea urchin D. mexicanum (up to 28.5 ind m⁻²) were recorded during monitoring, covering the areas where massive corals had been outplanted. Their bioerosive activity may have promoted partial tissue mortality and interrupted the calcification process, which is essential for coral attachment to the natural substrate (Herrera-Escalante et al., 2005; Omori, 2019). In addition, fragments were damaged by fish bites. Predation by reef fish is one of the main drivers of mortality for small coral recruits or, in this case, outplants of sizes \leq 5 cm2 (Ishida-Castañeda et al., 2020; Page et al., 2018; Palacios et al., 2014; Rivas et al., 2021), which decreases dramatically when fragments reach $\ge 25 \text{ cm}^2$ and attach to the substrate by tissue spreading (Tortolero-Langarica et al., 2020). Physical protection against coral predators increases survival and reduces predation effects (Hughes et al., 2007; Rivas et al., 2021), therefore it can be a potential tool to decrease mortality during the first growth phase, until outplants surpass a size that makes them less susceptible to predation and so increase their chances of long-term survival.

The lowest survivorship corresponds to P. lobata, which presents low dominance in the area, in contrast to coral reefs in the Central and South Pacific of Costa Rica (Cortés & Jiménez, 2003; Cortés et al., 2010; Jiménez, 2001). This might potentially indicate that local conditions at the restoration site are non-optimal for this species to prevail as a dominant reef-building coral. This situation contrasts with the high survivorship of Pocillopora spp. outplants, which used to be considered a highly sensitive genus to natural stressors (Glynn & Ault, 2000). However, they have recently proven to have the ability to acclimate to extreme conditions, especially thermal anomalies (Cruz-García et al., 2020; Romero-Torres et al., 2020) and upwelling conditions (Combillet et al., 2022). Due to their life history, it is expected that, in the short-term, Pocillopora spp. outplant survival will remain high, as colony size increases and competition for space and resources with other benthic fast-growing groups (e.g., macroalgae, turf) is reduced (Kodera et al., 2020; Lizcano-Sandoval et al., 2018; van Woesik et al., 2017).

Coral outplants in this study significantly increased their size in one year after their transplantation. Pocillopora spp. outplants experienced a 4-fold increase from their initial area in one year, while massive species showed a much slower growth: P. clavus and P. gigantea increased their initial area over 150 %, while P. lobata fragments did not even double their initial size. These differences between coral species respond to differences in their growth form, life history, local acclimatization, and reproduction strategies (Lirman & Schopmeyer, 2016; Page et al., 2018; Rinkevich, 2014). Pocillopora is a fast-growing genus, with growth rates over 5 cm yr⁻¹ (Tortolero-Langarica et al., 2017), and its branching morphology exponentially increases the three-dimensionality of the colony, which translates to a high contribution to coral cover and reef complexity (Kodera et al., 2020; Tortolero-Langarica et al., 2019). In contrast, slow growth rates of massive species result in a short-term low contribution to coral cover and reef heterogeneity (Forsman et al., 2015; Page et al., 2018; Rivas et al., 2021). Pocillopora outplants showed a high growth rate of 9.98 ± 1.69 cm yr⁻¹, in contrast to massive outplants, which not only showed low growth rates (0.61-1.48 cm yr⁻¹), but also experienced partial tissue mortality during monitoring period. The high effectiveness observed using Pocillopora fragments has been key for it to be the most widely used genus for restoration efforts throughout the ETP (Combillet et al., 2022; Ishida-Castañeda et al., 2020; Liñán-Cabello et al., 2011; Lizcano-Sandoval et al., 2018). Nonetheless, in order to achieve ecological rehabilitation, it is important to consider not only structural complexity but the calcification rate and calcium carbonate production, which is mostly provided by massive species (Tortolero-Langarica et al., 2022). Therefore, efforts should be focused on new approaches and protocols that would allow to increase the survival of the so-far challenging massive corals, and hence scale and maximize restoration efforts.

Although the rapid growth of *Pocillopora* is the result of its life history, and even more so in the study area, where growth rates are naturally higher than in other localities in the ETP (Jiménez & Cortés, 2003), other factors that promote the survival and success of this genus should be considered. Coral growth is influenced by local and site-specific conditions (Foo & Asner, 2020; Foo & Asner, 2021; Kodera et al., 2020). Bahía Culebra is influenced by seasonal upwelling, with sudden drops in seawater temperature down to 19 °C, which may elicit coral bleaching (Cupul-Magaña & Calderón-Aguilera, 2008; Lirman et al., 2011), and compromise growth and reproduction due to lack of energy available, but generally cause low mortality (Rodríguez-Troncoso et al., 2014). The ability of corals to maintain their costly physiological processes like growth and reproduction under such conditions is an acclimatization response (Gerstle, 2020; Rodríguez-Troncoso et al., 2010; Rodríguez-Troncoso et al., 2014; Rodríguez-Troncoso et al., 2016), as is the differential effect of abnormal high and low-temperature bleaching. Heat-induced bleaching causes an expulsion of the symbiont -its main energy supplier-, but also damages the polyp, while in abnormally low temperature conditions, bleaching results from photoinhibition of the symbiont (Lough & van Oppen, 2018; van Oppen & Blackall, 2019). However, the coral maintains its physiological processes by obtaining heterotrophic energy, and thus may help compensate its diminished autotrophic capacities (Jiménez & Cortés, 2003; Ziegler et al., 2014), which may cause a decrease but not inhibition of growth. Therefore, corals in Bahía Culebra must have the ability to acclimate to intense and sudden decreases in temperatures occurring during upwelling season. Moreover, unlike for massive corals, the presence of herbivores is beneficial, as D. mexicanum acts as a biological controller for the main coral competitors such as macroalgae and turf (Alvarado et al., 2012, Alvarado et al., 2016), thus decreasing competition pressure and also, generating space for possible sexual recruits (Cano et al., 2021; Graham et al., 2015; Roth et al., 2018).

The lower growth rate exhibited by *Pocillopora* outplants in the first month after their outplanting could be a result of what has been previously described as "transplantation shock". During this period, outplants undergo handling stress caused by the transplantation process (Precht, 2006), and acclimate to the environmental conditions of the new site; hence, their growth may be reduced (Afiq-Rosli et al., 2017; Forrester et al., 2012; Forrester et al., 2014; Lirman et al., 2010).

Our results show an influence of site of the donor colony on Pocillopora outplant growth. Differences in growth can be attributed to different genotypes and life history, even when outplanted to the same site (Drury et al., 2017; Goergen & Gilliam, 2018; Lirman et al., 2014; van Oppen et al., 2015). We do not possess information on every individual Pocillopora genotype outplanted to the reef; however, we hypothesize that the site of the donor colony could here be used as a proxy for different environmental tolerances, as fragments were originally collected from small areas in each donor site, and branching species reproduce mainly through fragmentation in the ETP (Bezy, 2009; Glynn et al., 2017; Sánchez-Noguera et al.; 2018b). Coral colonies from Matapalo site exhibited higher growth rates. Interestingly, this is the only site located outside the bay, is the most distant site from all others, and has relatively stable carbonate chemistry conditions (Sánchez-Noguera, 2019). Hence, the detected differences between donor sites when being subject to the same environmental conditions in the outplanting site could be a result of their life history and acclimation to the conditions of the donor site (Baums et al., 2019). Identifying growth differences between colonies and donor sites is relevant, even though trade-offs between growth, survival, resistance to disease, thermal tolerance and reproductive capacity may exist (Ware et al., 2020). Therefore, it is important to still outplant colonies with different life histories and responses to stress to the same restoration area, as this will maximize chances of survival in the face of changing conditions. Moreover, once outplants are sexually mature and reproduce among themselves, their offspring will likely receive considerable adaptive benefits (Baums et al., 2019), and hence ensure long-term success of restoration efforts.

As coral restoration is becoming a common strategy to recover and rehabilitate degraded coral reefs, it is essential to determine the best practices to ensure outplant survival and growth. These practices and techniques need to adapt to the local site-specific conditions for each area and outplanting site. Our results highlight the relevance of considering ecological processes such as herbivory and predation to maximize the success of restoration (Cano et al., 2021; Dang et al., 2020; Schopmeyer & Lirman, 2015) and how corals with different morphologies and life strategies may respond to the same environmental conditions. While Pocillopora spp. outplants had a positive response to outplanting technique and site conditions, fragments of massive species did experience partial tissue loss and dislodgement. Nevertheless, it is still important to restore coral reefs using a multi-species approach, since different coral species have different responses to disturbances (Lirman et al., 2011; Lustic et al., 2020). In the face of future changing conditions, promoting species richness and thus ecosystem resilience is key to ensure long-term recovery of coral reefs. Moreover, although monitoring individual outplants in the first year is vital to evaluate the feasibility of transplantation, there is a need to increase monitoring timeframes and develop appropriate ecological indicators in order to evaluate restoration effectiveness (Hein et al., 2017). These results could inform future practices and improve success of restoration efforts in the ETP.

Ethical statement: the authors declare that they all agree with this publication and made significant contributions; that there is no conflict of interest of any kind; and that we followed all pertinent ethical and legal procedures and requirements. All financial sources are fully and clearly stated in the acknowledgements section. A signed document has been filed in the journal archives.

ACKNOWLEDGMENTS

The present study would not have been possible without the support of Centro de Investigación en Ciencias del Mar y Limnología (CIMAR), from Universidad de Costa Rica. This project was funded by Vicerrectoría de Investigación of Universidad de Costa Rica through Project B9089, Península Papagayo through their environmental sustainability program, and the German Agency for International Cooperation (GIZ). We are especially thankful to Carlos Marenco and the staff of Marina Papagayo for their support during fieldwork. We thank Camila Valverde for her help processing photographs, and Alma Paola Rodríguez-Troncoso and Sergio Madrigal-Mora for their suggestions and review of the manuscript.

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REFERENCES

- Afiq-Rosli, L., Taira, D., Loke, H.X., Toh, T.C., Toh, K.B., Ng, C.S.L., Cabaitan, P.C., Chou, L.M., & Song, T. (2017). *In situ* nurseries enhance coral transplant growth in sedimented waters. *Marine Biology Research*, 13(8), 878–887.
- Alvarado, J. J., Beita-Jiménez, A., Mena, S., Fernández-García, C., Cortés, J., Sánchez-Noguera, C., Jiménez, C., & Guzmán-Mora, A. G. (2018). Cuando la conservación no puede seguir el ritmo del desarrollo: Estado de salud de los ecosistemas coralinos del Pacífico Norte de Costa Rica. *Revista de Biología Tropical*, 66(S1), 280–308.
- Alvarado, J. J., Cortés, J., Guzmán, H., & Reyes-Bonilla, H. (2016). Density, size, and biomass of *Diadema mexicanum* (Echinoidea) in Eastern Tropical Pacific coral reefs. *Aquatic Biology*, 24, 151–161.
- Alvarado, J. J., Cortés, J., & Reyes-Bonilla, H. (2012). Reconstruction of *Diadema mexicanum* bioerosion impact on three Costa Rican Pacific coral reefs. *Revista de Biología Tropical*, 60(S2), 121–132.
- Arnold, S. N., Steneck, R. S., & Mumby, P. J. (2010). Running the gauntlet: inhibitory effects of algal turfs on the processes of coral recruitment. *Marine Ecology Progress Series*, 414, 91–105.
- Baums, I. B., Baker, A. C., Davies, S. W., Grottoli, A. G., Kenkel, C. D., Kitchen, S. A., Kuffner, I. B., LaJeunesse, T. D., Matz, M. V., Miller, M. W., Parkinson, J. E., & Shantz, A. A. (2019). Considerations for maximizing the adaptive potential of restored coral populations in the western Atlantic. *Ecological Applications*, 29(8), e01978.
- Bayraktarov, E., Banaszak, A. T., Montoya-Maya, P., Kleypas, J., Arias-González, J. E., Blanco, M., Calle-Triviño, J., Charuvi, N., Cortés-Useche, C., Galván, V., García-Salgado, M. A., Gnecco, M., Guendulain-García, S. D., Hernández-Delgado, E. A., Marín-Moraga, J. A., Maya, M. F., Mendoza-Quiroz, S., Mercado-Cervantes, S., Morikawa, M., Nava, G. ...

Frías-Torres, S. (2020). Coral reef restoration efforts in Latin American countries and territories. *PLoS ONE*, *15* (8), e0228477.

- Bezy, M. B. (2009). Reproducción sexual y reclutamiento del coral masivo Pavona clavus en Bahía Culebra, Golfo de Papagayo, Costa Rica [Unpublished master's thesis]. Universidad de Costa Rica.
- Boström-Einarsson, L., Babcock, R. C., Bayraktarov, E., Ceccarelli, D., Cook, N., Ferse, S. C. A., Hancock, B., Harrison, P., Hein, M., Shaver, E., Smith, A., Suggett, D., Stewart-Sinclair, P. J., Vardi, T., & McLeod, I. M. (2020). Coral restoration – A systematic review of current methods, successes, failures and future directions. *PLoS ONE*, 15(1), e0226631.
- Cano, I., Sellares-Blasco, R. I., Lefcheck, J. S., Villalpando, M. F., & Croquer, A. (2021). Effects of herbivory by the urchin *Diadema antillarum* on early restoration success of the coral *Acropora cervicornis* in the central Caribbean. *Journal of Experimental Marine Biology and Ecology*, 539, 151541.
- Chomitz, B. (2021). The coral-associated fauna and sessile benthic reef community of Pocillopora coral colonies following transplantation in Golfo Dulce, South Pacific, Costa Rica: Composition, dynamics and succession [Unpublished master's thesis]. Universidad de Costa Rica.
- Coles, S. L., & Jokiel, P. L. (1978). Synergistic effects of temperature, salinity and light on the hermatypic coral *Montipora verrucosa*. *Marine Biology*, 49, 187–195.
- Combillet, L., Fabregat-Malé, S., Mena, S., Marín-Moraga, J. A., Gutiérrez, M., & Alvarado, J. J. (2022). *Pocillopora* spp. growth analysis on restoration structures in an Eastern Tropical Pacific upwelling area. *PeerJ*, *10*, e13248.
- Cortés, J., & Jiménez, C. (2003). Corals and coral reefs of the Pacific of Costa Rica: history, research and status. In J. Cortés (Ed.), *Latin American Coral Reefs* (pp. 361-367). Elsevier Science.
- Cortés, J., Jiménez, C. E., Fonseca, A. C, & Alvarado, J. J. (2010). Status and conservation of coral reefs in Costa Rica. *Revista de Biología Tropical*, 58(S1), 33–50.
- Cruz-García, R., Rodríguez-Troncoso, A. P., Rodríguez-Zaragoza, F. A., Mayfield, C. A. & Cupul-Magaña, A. L. (2020). Ephemeral effects of El Niño–Southern Oscillation events on an eastern tropical Pacific coral community. *Marine and Freshwater Research*, 71(10), 1259–1268.
- Cupul-Magaña, A. L., & Calderón-Aguilera, L. (2008). Cold water bleaching at Islas Marietas National Park, Nayarit, Mexico. XV Oceanography National Meeting, Veracruz, Mexico.

- D'Angelo, C., & Wiedenmann, J. (2014). Impacts of nutrient enrichment on coral reefs: new perspectives and implications for coastal management and reef survival. *Current Opinion in Environmental Sustainability*, 7, 82–93.
- Dang, V. D. H., Cheung, P. Y., Fong, C. L., Mulla, A. J., Shiu, J. H., Lin, C. H., & Nozawa, Y. (2020) Sea urchins play an increasingly important role for coral resilience across reefs in Taiwan. *Frontiers in Marine Science*, 7, 581945.
- Drury, C., Manzello, D. P., & Lirman, D. (2017). Genotype and local environment dynamically influence growth, disturbance response and survivorship in the threatened coral, *Acropora cervicornis*. *PLoS ONE*, *12*, e0174000.
- Edmunds, P. J., & Putnam, H. M. (2020). Science-based approach to using growth rate to assess coral performance and restoration outcomes. *Biology Letters*, 1, 20200227.
- Edwards, A. J. (2010). Reef rehabilitation manual. Coral reef targeted research & capacity building for Management Program. St Lucia, Australia.
- Faxneld, S., Jörgensen, T. L., & Tedengren, M. (2010). Effects of elevated seawater temperature, reduced salinity and nutrient enrichment on metabolism of the coral *Turbinaria mesenterina*. *Estuarine, Coastal and Shelf Science*, 88(4), 482–487.
- Fernández, C. (2007). Propagación del alga Caulerpa sertularioides (Chlorophyta) en Bahía Culebra, Golfo de Papagayo, Pacífico norte de Costa Rica [Unpublished master's thesis]. Universidad de Costa Rica.
- Fernández-García, C., Cortés, J., Alvarado, J. J., & Nívia-Ruiz, J. (2012). Physical factors contributing to the benthic dominance of the algae *Caulerpa sertularioides* (Caulerpaceae, Clorophyta) in the upwelling Bahía Culebra, North Pacific of Costa Rica. *Revista de Biología Tropical*, 60, 93–107.
- Ferse, S. C. A., Hein, M. Y., & Rölfer L. (2021). A survey of current trends and suggested future directions in coral transplantation for reef restoration. *PLoS ONE*, *16*(5), e0249966.
- Foo, S. A, & Asner, G. P. (2020). Sea surface temperature in coral reef restoration outcomes. *Environmental Research Letters*, 15, 074045.
- Foo, S. A., & Asner, G. P. (2021). Impacts of remotely sensed environmental drivers on coral outplant survival. *Restoration Ecology*, 29(1), e13309.
- Forrester, G. E., Ferguson, M. A., O'Connell-Rodwell, C. E., & Jarecki, L. L. (2014). Long-term survival and colony growth of *Acropora palmata* fragments transplanted by volunteers for restoration. *Aquatic Conservation: Marine and Freshwater Ecosystems*, 24, 81–91.

- Forrester, G. E., Maynard, A., Schofield, S., & Taylor, K. (2012). Evaluating causes of transplant stress in fragments of *Acropora palmata* used for coral reef restoration. *Bulletin of Marine Science*, 88(4), 1099–1113.
- Forsman, Z. H., Page, C. A., Toonen, R. J., & Vaughan, D. (2015). Growing coral larger and faster: microcolony-fusion as a strategy for accelerating coral cover. *PeerJ*, 3, e1313.
- Gerstle, L. (2020). Growth and transgenerational acclimatization of juvenile Pocillopora damicornis [Unpublished doctoral dissertation]. University of Hawai'i.
- Glynn, P., & Ault, J. (2000). A biogeographic analysis and review of the far eastern Pacific coral reef region. *Coral Reefs*, 19, 1–23.
- Glynn, P. W., Colley, S. B., Carpizo-Ituarte, E., & Richmond, R. H. (2017). Coral reproduction in the Eastern Pacific. In P. W. Glynn, D. P. Manzello, I. C. Enochs (Eds.), *Coral Reefs of the Eastern Tropical Pacific* (Vol 8, pp. 435–476). Springer Science+Business Media.
- Goergen, E. A., & Gilliam, D. S. (2018). Outplanting technique, host genotype, and site affect the initial success of outplanted *Acropora cervicornis*. *PeerJ*, *6*, e4433.
- Graham, N. A. J., Jennings, S., MacNeil, M. A., Mouillot, D., & Wilson, S. K. (2015). Predicting climate-driven regime shifts versus rebound potential in coral reefs. *Nature*, 518, 1–17.
- Guzmán, H. (1991). Restoration of coral reefs in Pacific Costa Rica. *Conservation Biology*, 5(2), 189–195.
- Hein, M. Y., Willis, B. E., Beeden, R., & Birtles, A. (2017). The need for broader ecological and socioeconomic tools to evaluate the effectiveness of coral restoration programs. *Restoration Ecology*, 25(6), 873–883.
- Herrera-Escalante, T., López-Pérez, R. A., & Leyte-Morales, G. E. (2005). Bioerosion caused by the sea urchin *Diadema mexicanum* (Echinodermata: Echinoidea) at Bahías de Huatulco, Western Mexico. *Revista de Biología Tropical*, 53(S3), 263–273.
- Hughes, T. P., Kerry, J., Álvarez-Noriega, M., Álvarez-Romero, J. G., Anderson, K. D., Baird, A. H., Babcock, R. C., Beger, M., Bellwood, D. R., Berkelmans, R., Bridge, T. C., Butler, I. R., Byrne, M., Cantin, N. E., Comeau, S., Connolly, S. R., Cumming, G. S., Dalton, S. J., Diaz-Pulido, G., ... Wilson, S. K. (2017). Global warming and recurrent mass bleaching of corals. *Nature*, 543, 373–377.
- Hughes, T. P., Kerry, J. T., Baird, A. H. Connolly, S. R., Chase, T. J., Dietzel, A., Hill, T., Hoey, A. S., Hoogenboom, M. O., Jacobson, M., Kerswell, A., Madin, J. S., Mieog, A., Paley, A. S., Pratchett, M. S., Torda, G., Woods, R. M. (2019). Global warming impairs

stock-recruitment dynamics of corals. *Nature*, 568, 387-390.

- Hughes, T. P., Rodrigues, M. J., Bellwood, D. R., Ceccarelli, D., Hoegh-Guldberg, O., McCook, L., Moltschaniwskyj, N., Pratchett, M. S., Steneck, R. S., & Willis, B. (2007). Phase shifts, herbivory, and the resilience of coral reefs to climate change. *Current Biology*, 17(4), 360–365.
- Ishida-Castañeda, J., Pizarro, V., López-Victoria, M., & Zapata, F. A. (2020). Coral reef restoration in the Eastern Tropical Pacific: feasibility of the coral nursery approach. *Restoration Ecology*, 28(1), 22–28.
- Jiménez, C. (1997). Corals and coral reefs of Culebra Bay, Pacific coast of Costa Rica: anarchy in the reef [Paper presentation]. In H.A. Lessios & I.A. Macintyre (Eds.), Proceedings of the VIII International Coral Reef Symposium [Symposium], VIII International Coral Reef Symposium, Panama.
- Jiménez, C. (2001). Arrecifes y ambientes coralinos de Bahía Culebra, Pacífico de Costa Rica: aspectos biológicos, económicos, recreativos y de manejo. *Revista de Biología Tropical*, 49(S2), 215–231.
- Jiménez, C. (2007). Arrecifes coralinos, ¿víctimas de los cambios? *Ambientico*, 171, 5–7.
- Jiménez, C., Bassey, G., Segura, Á., & Cortés, J. (2010). Characterization of the coral communities and reefs of two previously undescribed locations in the upwelling region of Gulf of Papagayo (Costa Rica). *Revista Ciencias Marinas y Costeras*, 2, 95–108.
- Jiménez, C., & Cortés, J. (2003). Growth of seven species of scleractinian corals in an upwelling environment of the Eastern Pacific (Golfo de Papagayo, Costa Rica). *Bulletin of Marine Science*, 72, 187–198.
- Kleypas, J. A., & Yates, K. K. (2009). Coral reefs and ocean acidification. *Oceanography*, 22(4), 108–117.
- Kleypas, J. A., McManus, J. W., & Meñez L. A. B. (1999). Environmental limits to coral reef development: Where do we draw the line? *American Zoologist*, 39(1), 146–159.
- Kodera, S. M., Edwards, C. B., Petrovic, V., Pedersen, N. E., Eynaud, Y., & Sandin, S. A. (2020). Quantifying life history demographics of the scleractinian coral genus *Pocillopora* at Palmyra Atoll. *Coral Reefs*, 39, 1091–1105.
- Koval, G., Rivas, N., D'Alessandro, M., Hesley, D., Santos, R., & Lirman, D. (2020). Fish predation hinders the success of coral restoration efforts using fragmented massive corals. *PeerJ*, 8, e9978.
- Ladd, M. C., Miller, M. W., Hunt, J. H., Sharp, W. C., & Burkpile, D. E. (2019). Harnessing ecological processes to facilitate coral restoration. *Frontiers in Ecology* and the Environment, 16(4), 239–247.

- Liñán-Cabello, M. A., Flores-Ramírez, L. A., Laurel-Sandoval, M. A., García-Mendoza, E., Soriano-Santiago, O., & Delgadillo-Nuño, M. A. (2011). Acclimation in *Pocillopora* spp. during a coral restoration program in Carrizales Bay, Colima, Mexico. *Marine and Freshwater Behaviour and Physiology*, 44(1), 61–72.
- Lirman, D., & Schopmeyer, S. (2016). Ecological solutions to reef degradation: optimizing coral reef restoration in the Caribbean and Western Atlantic. *PeerJ*, 4, e2597.
- Lirman, D., Schopmeyer, S., Galvan, V., Drury, C., Baker, A. C., & Baums, I. B. (2014). Growth dynamics of the threatened Caribbean staghorn coral *Acropora cervicornis*: influence of host genotype, symbiont identity, colony size, and environmental setting. *PLoS ONE*, 9(9), e107253.
- Lirman, D., Schopmeyer, S., Manzello, D., Gramer, L. J., Precht, W. F., Muller-Karger, F., Banks, K., Barnes, B., Bartels, E., Bourque, A., Byrne, J., Donahue, S., Duquesnel, J., Fisher, L., Gilliam, D., Hendee, J., Johnson, M., Maxwell, K., McDevitt, E., Monty, J., Rueda, D., Ruzicka, R., & Thanner, S. (2011). Severe 2010 cold-water event caused unprecedented mortality to corals of the Florida reef tract and reversed previous survivorship patterns. *PLoS ONE*, *6*(8), e23047.
- Lirman, D., Thyberg, T., Herlan, J., Hill, C., Young-Lahiff, C., Schopmeyer, S., Huntington, B., Santos, R., & Drury, C. (2010). Propagation of the threatened staghorn coral *Acropora cervicornis*: methods to minimize the impacts of fragment collection and maximize production. *Coral Reefs*, 29, 729–735.
- Lizcano-Sandoval, L. D., Londoño-Cruz, E., & Zapata, F. A. (2018). Growth and survival of *Pocillopora damicornis* (Scleractinia: Pocilloporidae) coral fragments and their potential for reef restoration in the Tropical Eastern Pacific. *Marine Biology Research*, 14, 887–897.
- Lohr, K. E., Ripple, K., & Patterson, J. T. (2020). Differential disturbance effects and phenotypic plasticity among outplanted corals at patch and fore reef sites. *Journal for Nature Conservation*, 55, 125827.
- Lough, J. M., & van Oppen, M. J. H. (2018). Introduction: coral bleaching-patterns, processes, causes and consequences. In M. J. H. van Oppen, & J. M. Lough (Eds.), *Coral Bleaching, Patterns, processes, causes,* and consequences (pp. 1–8). Springer.
- Lustic, C., Maxwell, K., Bartels, E., Reckenbeil, B., Utset, E., Schopmeyer, S., Zink, I., & Lirman, D. (2020). The impacts of competitive interactions on coral colonies after transplantation: A multispecies experiment from the Florida Keys, US. *Bulletin of Marine Science*, 96(4), 805–818.
- Miller, M. W., Lohr, K. E., Cameron, C. M., Williams, D. E., & Peters, E. C. (2014). Disease dynamics

and potential mitigation among restored and wild staghorn coral, Acropora cervicornis. PeerJ, 2, e541.

 \odot

- Omori, M. (2019). Coral restoration research and technical developments: what we have learned so far. *Marine Biology Research*, 15(7), 377–409.
- Page, C. A., Muller, E. M., & Vaughan, D. E. (2018). Microfragmenting for the successful restoration of slow growing massive corals. *Ecological Engineering*, 123, 86–94.
- Palacios, M. M., Muñoz, C. G., & Zapata, F. A. (2014). Fish corallivory on a pocilloporid reef and experimental coral responses to predation. *Coral Reefs*, 33, 625–636.
- Pinzón, J. H., & LaJeunesse, T. C. (2011). Species delimitation of common reef corals in the genus *Pocillopora* using nucleotide sequence phylogenies, population genetics and symbiosis ecology. *Molecular Ecology*, 20(2), 311–325.
- Pinzón, J. H., Sampayo, E., Cox, E., Chauka, L. J., Chen, C. A., Voolstra, C. R., & LaJeunesse, T. C. (2013). Blind to morphology: genetics identifies several widespread ecologically common species and few endemics among Indo-Pacific cauliflower corals (*Pocillopora*, Scleractinia). *Journal of Biogeography*, 40(8), 1595–1608.
- Precht, W. F. (2006). Coral reef restoration handbook: the rehabilitation of an ecosystem under siege. CRC Press.
- R Core Team (2021). R: A language and environment for statistical computing [Computer software]. R Foundation for Statistical Computing. http://www.rproject.org/index.html
- Rinkevich, B. (1995). Restoration strategies for coral reefs damaged by recreational activities: The use of sexual and asexual recruits. *Restoration Ecology*, 3(4), 241–251.
- Rinkevich, B. (2014). Rebuilding coral reefs: does active reef restoration lead to sustainable reefs? *Current Opinion in Environmental Sustainability*, 7, 28–36.
- Rivas, N., Hesley, D., Kaufman, M., Unsworth, J., D'Alessandro, M., & Lirman, D. (2021). Developing best practices for the restoration of massive corals and the mitigation of predation impacts: influences of physical protection, colony size, and genotype on outplant mortality. *Coral Reefs*, 40, 1227–1241.
- Rixen, T., Jiménez, C., & Cortés, C. (2012). Impact of upwelling events on the sea water carbonate chemistry and dissolved oxygen concentration in the Gulf of Papagayo (Culebra Bay), Costa Rica: Implications for coral reefs. *Revista de Biología Tropical*, 60(S2), 187–195.

- Rodríguez-Sáenz, K., & Rodríguez-Fonseca, J. (2004). Avistamientos del delfín manchado, *Stenella attenuata* (Cetacea: Delphinidae) en Bahía Culebra, Costa Rica, 1999-2000. *Revista de Biología Tropical*, 49(2), 189–193.
- Rodríguez-Troncoso, A. P., Carpizo-Ituarte, E., & Cupul-Magaña, A. L. (2010). Differential response to cold and warm water conditions in *Pocillopora* colonies from the Central Mexican Pacific. *Journal of Experimental Marine Biology and Ecology*, 391(1–2), 57–64.
- Rodríguez-Troncoso, A. P., Carpizo-Ituarte, E., & Cupul-Magaña, A. L. (2016). Physiological response to high temperature in the Tropical Eastern Pacific coral *Pocillopora verrucosa*. *Marine Ecology*, 37(5), 1168–1175.
- Rodríguez-Troncoso, A. P., Carpizo-Ituarte, E., Pettay, D. T., Warner, M. E., & Cupul-Magaña, A. L. (2014). The effects of an abnormal decrease in temperature on the Eastern Pacific reef-building coral *Pocillopora verrucosa*. *Marine Biology*, *161*, 131–139.
- Romero-Torres, M., Acosta, A., Palacio-Castro, A. M., Treml, E. A., Zapata, F. A., Paz-García, D. A., & Porter, J. W. (2020). Coral reef resilience to thermal stress in the Eastern Tropical Pacific. *Global Change Biology*, 26(7), 3880–3890.
- Roth, F., Saalmann, F., Thomson, T., Coker, D. J., Villalobos, R., Jones, B. H., Wild, C., & Carvalho, S. (2018). Coral reef degradation affects the potential for reef recovery after disturbance. *Marine Environmental Research*, 142, 48–58.
- Sánchez-Noguera, C. (2012). Cambios socioeconómicos y ambientales en Bahía Culebra, Guanacaste, Costa Rica: implicaciones para su gestión [Unpublished master's tesis]. Universidad de Costa Rica.
- Sánchez-Noguera, C. (2019). Carbonate chemistry and coral reefs in the Pacific coast of Costa Rica [Unpublished doctoral dissertation]. Universität Hamburg.
- Sánchez-Noguera, C., Jiménez, C., & Cortés, J. (2018a). Desarrollo costero y ambientes marino-costeros en Bahía Culebra, Guanacaste, Costa Rica. *Revista de Biología Tropical*, 66(S1), 309–327.
- Sánchez-Noguera, C., Stuhldreier, I., Cortés, J., Jiménez, C., Morales, Á., Wild, C., & Rixen, T. (2018b). Natural ocean acidification at Papagayo upwelling system (north Pacific Costa Rica): implications for reef development. *Biogeosciences*, 15, 2349–2360.
- Schopmeyer, S. A., & Lirman, D. (2015). Occupation dynamics and impacts of damselfish territoriality on recovering populations of the threatened staghorn coral, *Acropora cervicornis. PLoS ONE*, 10(11), e0141302.
- Shafir, S., & Rinkevich, B. (2013). Mariculture of coral colonies for the public aquarium sector. In R. J.

Leewis, & M. Janse (Eds.), *Advances in coral husbandry in public aquariums* (Vol 2, pp. 315–318). Burgers' Zoo.

- Stuhldreier, I., Sánchez-Noguera, C., Roth, C., Cortés, J., Rixen, T., & Wild, C. (2015). Upwelling increases net primary production of corals and reef-wide gross primary production along the Pacific coast of Costa Rica. *Frontiers in Marine Science*, 2, 113.
- Tortolero-Langarica, J. J. A., Cupul-Magaña, A. L., & Rodríguez-Troncoso, A. P. (2014). Restoration of a degraded coral reef using a natural remediation process: A case study from a Central Mexican Pacific National Park. Ocean & Coastal Management, 96, 12–19.
- Tortolero-Langarica, J. J. A, Rodríguez-Troncoso, A. P., Cupul-Magaña, A. L., & Carricart-Ganivet, J. P. (2017). Calcification and growth rate recovery of the reef-building *Pocillopora* species in the northeast tropical Pacific following an ENSO disturbance. *PeerJ*, 5, e3191.
- Tortolero-Langarica, J. J. A., Rodríguez-Troncoso, A. P., Cupul-Magaña, A. L., Alarcón-Ortega, L. C., Santiago-Valentín, J. D. (2019). Accelerated recovery of calcium carbonate production in coral reefs using low-tech ecological restoration. *Ecological Engineering*, 128, 89–97.
- Tortolero-Langarica, J. J. A., Rodríguez-Troncoso, A. P., Cupul-Magaña, A. L., & Rinkevich, B. (2020). Micro-fragmentation as an effective and applied tool to restore remote reefs in the Eastern Tropical Pacific. *International Journal of Environmental Research and Public Health*, 17, 6574.
- Tortolero-Langarica, J. J. A., Rodríguez-Troncoso, A. P., Cupul-Magaña, A. L., Morales-de-Anda, D. E., Caselle, J. E., & Carricart-Ganivet, J. P. (2022). Coral calcification and carbonate production in the eastern tropical Pacific: The role of branching and massive corals in the reef maintenance. *Geobiology*, 20(4), 533–545.
- Trygonis, V., & Sini, M. (2012). PhotoQuad: A dedicated seabed image processing software, and a comparative error analysis of four photoquadrat methods. *Journal of Experimental Marine Biology and Ecology*, 424–425, 99–108.
- Unsworth, J. D., Hesley, D., D'Alessandro, M., & Lirman, D. (2020). Outplanting optimized: Developing a more efficient coral attachment technique using Portland cement. *Restoration Ecology*, 29(1), e13299.
- van Oppen, M. J. H., & Blackall, L. L. (2019). Coral microbiome dynamics, functions and design in a changing world. *Nature Reviews Microbiology*, 17, 557–567.
- van Oppen, M. J. H., Oliver, J. K., Putnam, H. M., & Gates, R. D. (2015). Building coral reef resilience through



assisted evolution. Proceedings of the National Academy of Sciences, 112, 2307–2313.

- van Woesik, R., Ripple, K., & Miller, S. L. (2017). Macroalgae reduces survival of nursery-reared *Acropora* corals in the Florida reef tract. *Restoration Ecology*, 26(3), 563–569.
- Ware, M., Garfield, E. N., Nedimyer, K., Levy, J., Kaufman, L., Precht, W., Winters, R. S., & Miller, S. L. (2020). Survivorship and growth in staghorn coral (*Acropora cervicornis*) outplanting projects in the

Florida Keys National Marine Sanctuary. *PLos ONE*, *15*(5), e0231817.

- Wiedenmann, J., D'Angelo, C., Smith, E.G., Hunt, A. N., Legiret, F. E., Postle, A. D., & Achterberg, E. P. (2013). Nutrient enrichment can increase the susceptibility of reef corals to bleaching. *Nature Climate Change*, *3*, 160–164.
- Ziegler, M., Roder, C. M., Büchel, C., & Voolstra, C. R. (2014). Limits to physiological plasticity of the coral *Pocillopora verrucosa* from the central Red Sea. *Coral Reefs*, 33, 1115–1129.